

# WORKFLOW DURING THE B3 CAMP

# MAKING SOLUTIONS FROM STOCKS

B3 Summer Science Camp  
at Olympic High School

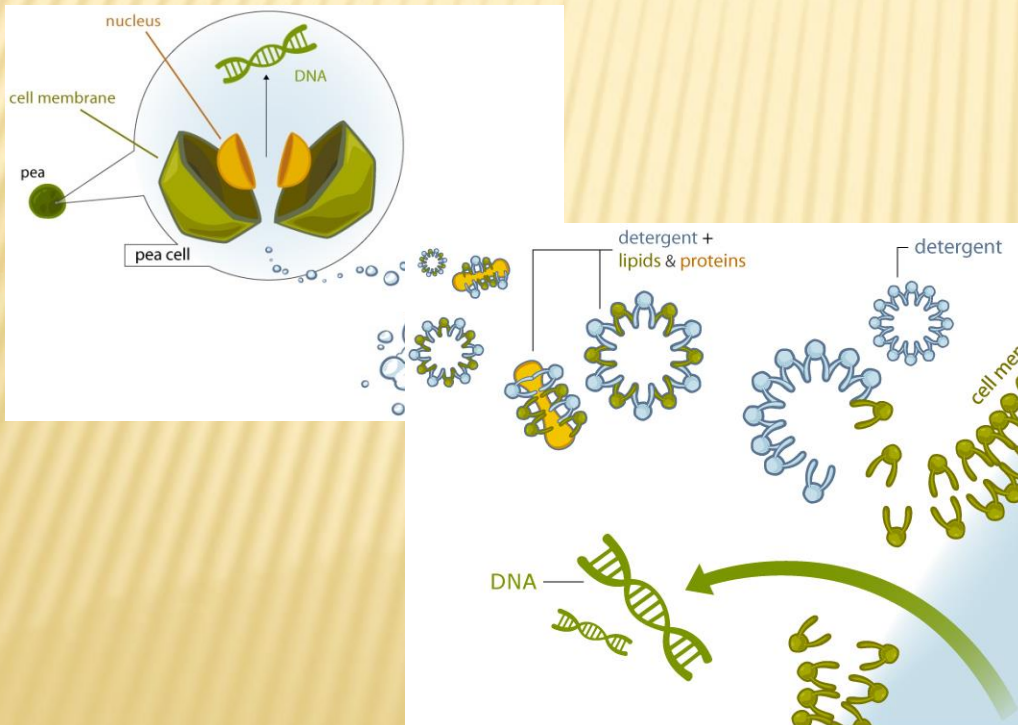


# LAB WORKFLOW OVERVIEW

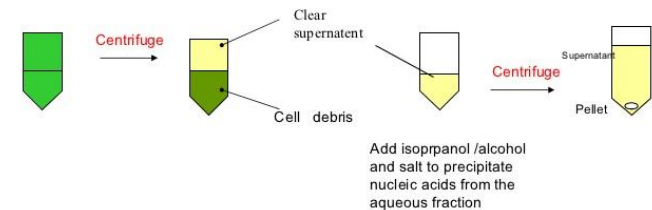
- Collect Samples (June 12<sup>th</sup>)
- Extract DNA from the samples (June 17<sup>th</sup>)
  - Break open the cells (contents will mix)
  - Use chemistry to extract nucleic acid from other cell contents and prevent degradation
  - Use biochemistry and chemistry to degrade the RNA and then concentrate the DNA
- Test the DNA for quality (June 18<sup>th</sup> and June 22<sup>nd</sup>)
  - How long are the pieces?
  - How much do we have?
  - How pure is it?
- Carry out two types of enzymatic reactions (June 23<sup>rd</sup> )
  - PCR for chloroplast segment, and agarose gel
  - Restriction Endonuclease (RE) digestion and size selection
- Construct a sequencing library with the RE material (June 24-25<sup>th</sup>)
  - Size select
  - Add small DNA ‘adaptors’ (with barcodes to tell apart) using DNA ligase
  - Carry out more PCR
  - Clean up the product to remove unwanted or unused components
- Test the library and quantify it (June 26<sup>th</sup>)
  - Verify the average length and concentration
  - Make the proper dilutions and pool the samples
- Sequence the pooled libraries (June 26<sup>th</sup>)

# EXTRACT DNA

- Extract DNA from the samples (June 17<sup>th</sup>)
  - Break open the cells (contents will mix)
  - Use chemistry to extract nucleic acid from other cell contents and prevent degradation
  - Use biochemistry and chemistry to degrade the RNA and then concentrate the DNA



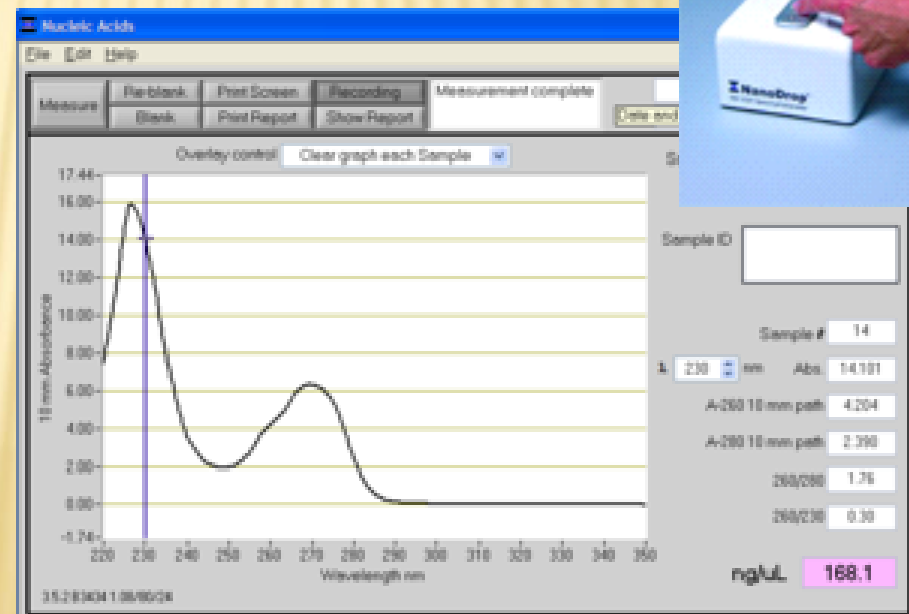
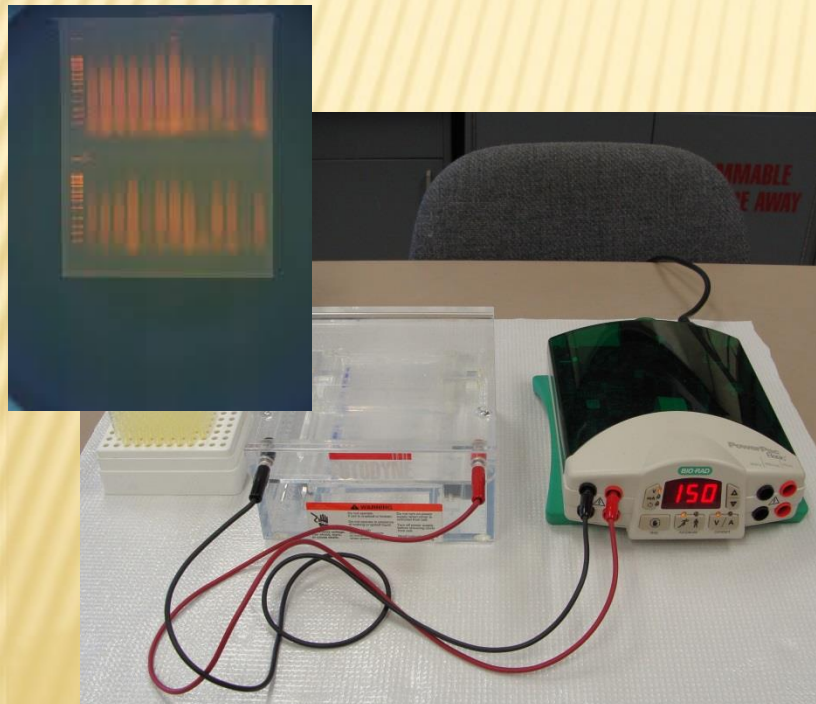
## Non-Phenol Chloroform based extraction of DNA for PCR detection of Citrus yellow mosaic virus and Citrus greening bacterium



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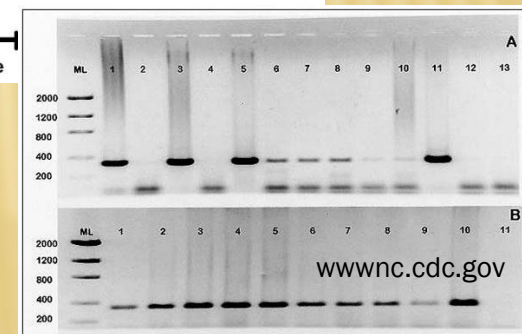
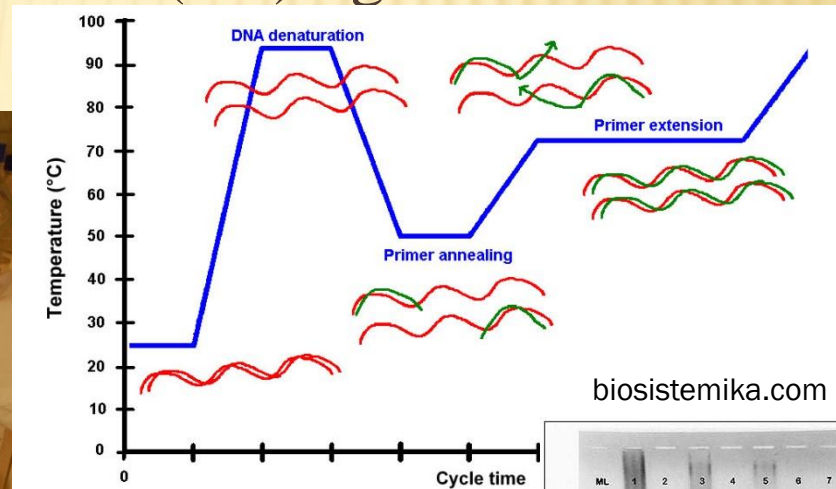
# QUALITY TESTING DNA

- Test the DNA for quality (June 18<sup>th</sup> and June 22<sup>nd</sup>)
  - How long are the pieces?
  - How much do we have? How pure is it?



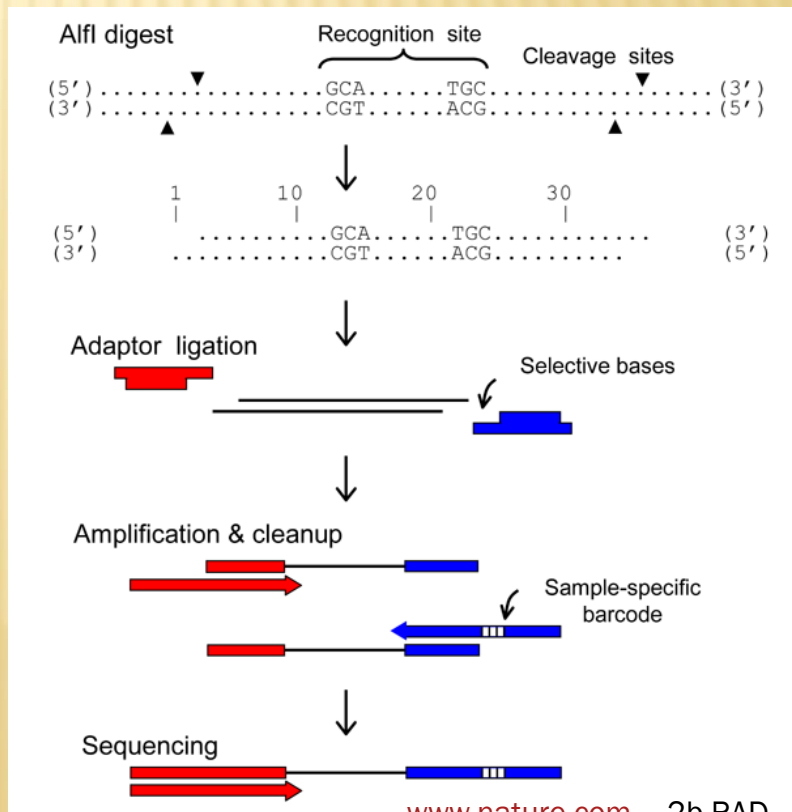
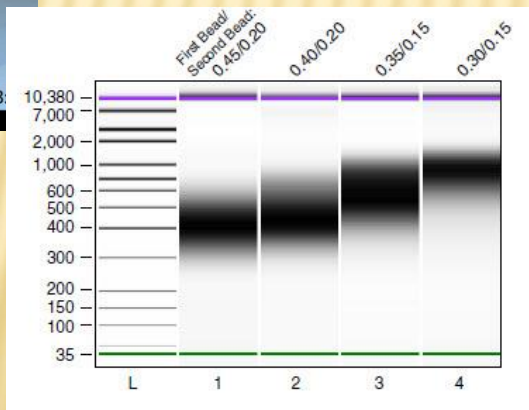
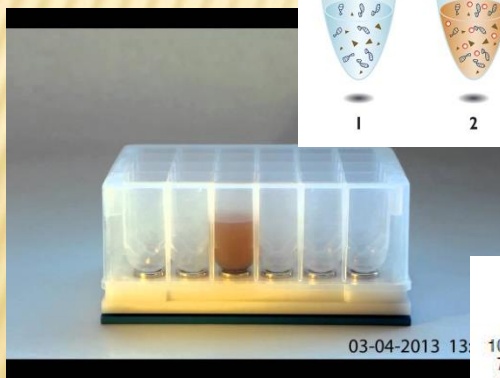
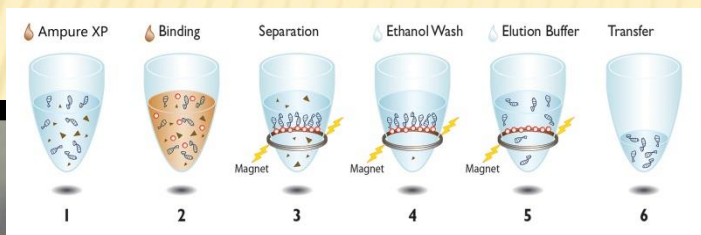
# PCR, RESTRICTION DIGESTION, LIGATION

- Carry out two types of enzymatic reactions (June 23<sup>rd</sup> )
  - PCR for chloroplast segment, and agarose gel
  - Restriction Endonuclease (RE) digestion and size selection



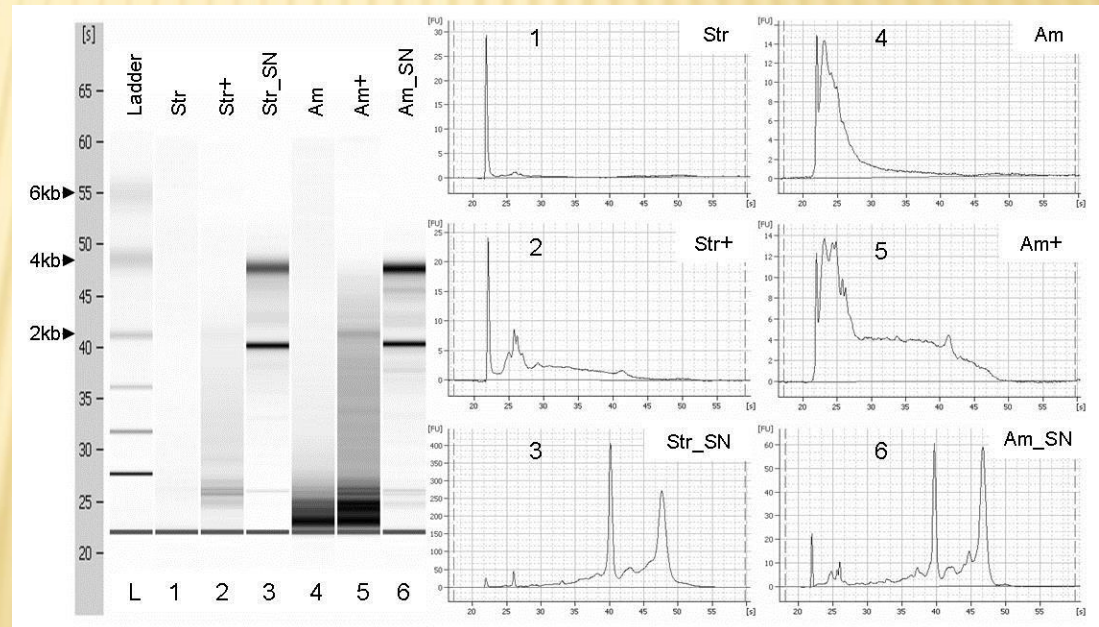
# SEQUENCING LIBRARY CONSTRUCTION

- Construct a sequencing library with the RE material (June 24-25<sup>th</sup>)
  - Size select with magnetic beads
  - Add small DNA ‘adaptors’ (with barcodes to tell apart) using DNA ligase
  - Use beads to clean up the product



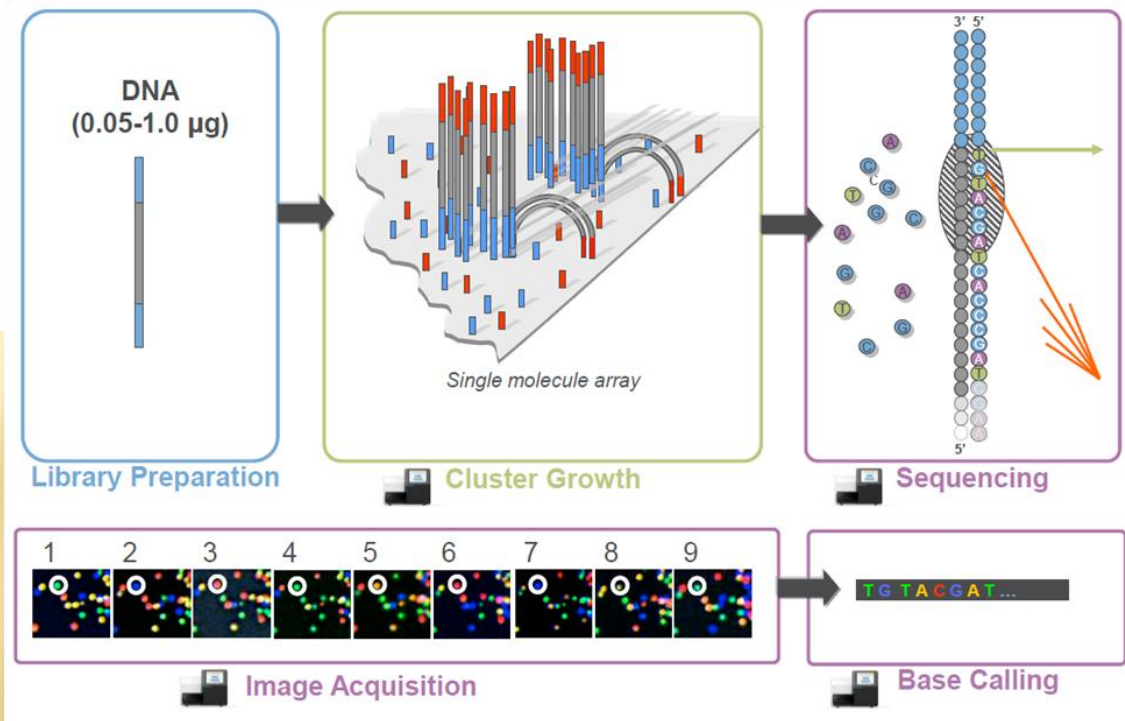
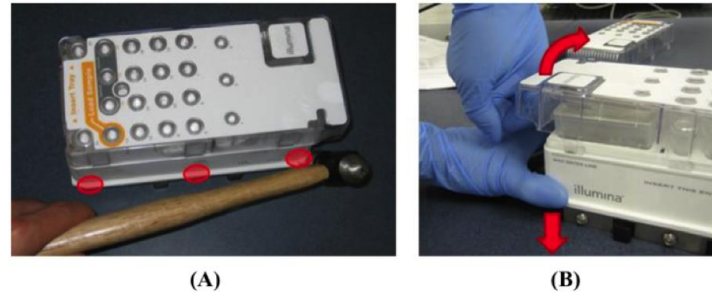
# SEQUENCING LIBRARY QUALITY TESTS

- Test the library and quantify it (June 26<sup>th</sup>)
  - Verify the average length and concentration
  - Make the proper dilutions and pool the samples



# SEQUENCING THE LIBRARY

- Sequence the pooled libraries (June 26<sup>th</sup>)





# LAB WORKFLOW - TODAY

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- Can you make solutions from stocks?
- Can you use microbalances properly?
- Do you know how to use micropipetters?

# LAB WORKFLOW - TODAY

- Can you make solutions from stocks?
  - Do you understand what different types of stocks are?



# LAB STOCKS – ‘FOLD’ STOCKS

- What is a 10X stock? How do you use it?



10-times more concentrated than standard use.

Take 1 part of the stock and add 9 parts of the other components.

If you are only adding water and you want 100ml (0.1L) of 1X TAE buffer

$$C_1V_1 = C_2V_2$$

$$(1X) 100\text{ml} = (10X) ?\text{ml}$$

$$[(1X) 100\text{ml} / 10X] = 10\text{ml}$$

For the rest of the solution: subtract 10 from 100 → 90ml of H<sub>2</sub>O if that is the only other component.

Result will be 100 ml of 1X TAE buffer.

# LAB STOCKS – ‘MOLAR’ STOCKS

- What is a 1.0 Molar stock?



Contains 1 mole of Tris molecules for every liter for solvent (Water in this case).

1 mole is Avogadro's number:  $6.023 \times 10^{23}$ .

Tris has a mass of 157.56 gm/mol so weigh out 157.56 grams, bring up to 1L with H<sub>2</sub>O (adjust the pH also).

If I want 100ml (0.1L) of a 50mM solution (0.05M), use the formula

$$C_1V_1 = C_2V_2$$

$$(0.05\text{M}) 0.1\text{L} = (1\text{M})X\text{L}$$

$$[(0.05\text{M}) 0.1\text{L}/1\text{M}] = 0.005\text{L} \text{ or } 5\text{ml}$$

So I will use 5ml of the stock and bring it to 100ml with the other components, which might be just water (so add 95ml) or several other things.

# LAB STOCKS – ‘MASS PER VOLUME’ STOCKS

- What is a Mass/Volume stock?



Contains 50 milligrams of the protein lysozyme (0.05 gm) per 1 ml (0.001 L) of solution.

This does not tell you what the solution contains – for proteins it is most likely some buffer like Tris at pH that stabilizes the enzyme, in water, and it might have some salt like  $\text{Ca}^{++}$  in it, and a small amount of glycerol (5% is common).

If I am making a 100 microliter (0.1 ml or  $100 \times 10^{-6} \text{L} = 10^{-4} \text{L}$ ) solution and I want it to have 1 mg/ml of lysozyme.

$$C_1V_1 = C_2V_2$$

$$1 \text{ mg/ml}(0.1 \text{ ml}) = 50 \text{ mg/ml}(X \text{ ml})$$

$$[1 \text{ mg/ml}(0.1 \text{ ml}) / 50 \text{ mg/ml}] = 0.002 \text{ ml or } 2 \text{ ul}$$

So I will use 2 ul of the stock and bring it to 100 ul with the other components to end up with 1 mg/ml of the lysozyme.

# LAB STOCKS – ‘PERCENT’ STOCKS

- What is a Percent stock?



Contains 10grams of the chemical SDS (sodium dodecyl sulfate, a detergent) per 100ml of solution (in this case the solvent is water – you have to read the chemical information sheet to know that).

Note: In molecular biology, when the main solvent (that dissolves or otherwise carries the other compounds) is not named it is assumed to be water.

If I am making a 100microliter (0.1ml or  $100 \times 10^{-6} \text{L} = 10^{-4} \text{L}$ ) solution and I want it to have a final concentration of 0.1% SDS:

$$C1V1 = C2V2$$

$$1\% (0.1 \text{ml}) = 10\% (X \text{ml})$$

$$[1\% (0.1 \text{ml}) / 10\%] = 0.010 \text{ml} \text{ or } 10 \text{ul}$$

So I will use 10ul of the stock and bring it to 100ul with the other components to end up with 1% SDS.

# LAB STOCKS – ‘UNIT’ STOCKS

- What is a Unit/Volume stock?



Contains 10grams of the chemical SDS (sodium dodecyl sulfate, a detergent) per 100ml of solution (in this case the solvent is water – you have to read the chemical information sheet to know that).

Note: In molecular biology, when the main solvent (that dissolves or otherwise carries the other compounds) is not named it is assumed to be water.

If I am making a 100microliter (0.1ml or  $100 \times 10^{-6} \text{L} = 10^{-4} \text{L}$ ) solution and I want it to have a final concentration of 0.1% SDS:

$$C1V1 = C2V2$$

$$1\% (0.1 \text{ml}) = 10\% (X \text{ml})$$

$$[1\% (0.1 \text{ml}) / 10\%] = 0.010 \text{ml} \text{ or } 10 \text{ul}$$

So I will use 10ul of the stock and bring it to 100ul with the other components to end up with 1% SDS.

# LAB STOCKS – USING CORRECTLY

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- Many protocols ask you to combine concentrated stocks with some of each kind listed above.
- The Using Lab Stocks Handout has examples of doing the calculations for a couple of the types of molecular biology protocols we will be using.